Photomodification of Blood Using Low-Intensity Optical Radiation

Photomodification of Blood Using Low-Intensity Optical Radiation

Ву

Galina Zalesskaya and Nalalya Mitkovskaya

Cambridge Scholars Publishing



Photomodification of Blood Using Low-Intensity Optical Radiation

By Galina Zalesskaya and Nalalya Mitkovskaya

This book first published 2020

Cambridge Scholars Publishing

Lady Stephenson Library, Newcastle upon Tyne, NE6 2PA, UK

British Library Cataloguing in Publication Data A catalogue record for this book is available from the British Library

Copyright © 2020 by Galina Zalesskaya and Nalalya Mitkovskaya

All rights for this book reserved. No part of this book may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission of the copyright owner.

ISBN (10): 1-5275-5861-4 ISBN (13): 978-1-5275-5861-8

TABLE OF CONTENTS

List of Figuresviii
List of Tablesxx
Acknowledgementsxxi
Introductionxxii
About the Structure of the Bookxxiv
Abbreviationsxxvii
Chapter One1
Effect of low-intensity optical radiation on the spectral-luminescent characteristics of blood irradiated <i>in vivo</i>
1.1 Electronic absorption spectra of whole blood, erythrocytes and plasma
1.2 Infrared absorption spectra of whole blood, erythrocytes and
plasma
1.4 Plasma fluorescence 14
1.5 Spectral manifestation of blood photomodification
and plasma under UV phototherapy
1.5-3 The effect of laser phototherapy at wavelengths of 632.8 nm and 780 nm on infrared absorption spectra of blood,
erythrocytes and plasma
under UV phototherapy
1.7 References

Chapter Two
Photomodification of blood irradiated <i>in vivo</i> using low-intensity optical
radiation
2.1 Gas composition of venous blood under phototherapy
2.2 The effect of phototherapy on hemoglobin oxygen saturation 58
2.3 Hemoglobin-oxygen affinity and oxygen capacity of blood under
the phototherapy62
2.4 The effect of phototherapy on erythrocytes of blood
2.4-1 Changes in the shape of red blood cells under the
phototherapy71
2.4-2 The electrophoretic mobility of erythrocytes under UVBI
and magnetic therapy
2.4-3 The effect of phototherapy on the erythrocyte metabolism 78
2.5. The individualization of therapeutic doses of optical radiation
according to blood oxygenation
2.6 Photomodification of blood aggregation characteristics under
phototherapy85
2.7 The phototherapy effect on glucose and lactate concentrations 92
2.8 The correction of lipid metabolism by phototherapy
2.8-1 The effect of UV blood irradiation on lipid metabolism 97
2.8-2 The effect of overvein blood irradiation on lipid
metabolism
2.9 References
ort
Chapter Three
Molecular mechanisms of the phototherapy action
3.1 Primary photoacceptors and primary photoreactions (literature
data)
3.2 Interaction of optical radiation with human tissues under
the phototherapy
3.2-1 Optical characteristics of blood and skin tissues,
and the penetration depth of optical radiation
3.2-2 Blood photomodification by optical radiation at various
wavelengths
3.2-3 The heating of human tissue during phototherapy: computer
simulation
infrared laser radiation
3.2-5 Modeling of laser irradiation conditions for mucosal tissues
in an antimicrobial photodynamic therapy
3.3 The effect of magnetic therapy on blood molecules

3.4 The molecular mechanism of the phototherapy action based	
on the absorption of optical radiation by hemoglobin	2
3.5 The role of the reactive oxygen species in the therapeutic action	
of low-intensity optical radiation	5
3.5-1 Destruction of human tissue biomolecules initiated	
by reactive oxygen species	5
3.5-2 Positive effects of reactive oxygen species during	
the phototherapy160	6
3.6 References	1
Chapter Four	
The combined action of gamma radiation and low-intensity laser radiation	
on experimental animals (rats)	
4.1 The radioprotective action of laser radiation (literature data) 180	0
4.2 The combined effect of gamma and low-intensity laser radiation	
on peripheral blood of rats: pre- and post- gamma exposure	
of blood	
4.2-1 Goal of studies; materials and methods	6
4.2-2 Electronic absorption spectra of blood at combined	
irradiation	9
4.2-3 The effect of gamma radiation on leucocyte (WBC),	
lymphocyte (LYM) and granulocyte (GRA) in peripheral	_
blood	2
4.2-4 The effect of gamma radiation on erythrocyte (RBC)	_
and hemoglobin concentrations, hematocrit (Hct) values 195	5
4.2-5 The effect of gamma radiation on platelet (PLT)	_
concentration	7
4.2-6 The effect of gamma radiation on activity of antioxidant	_
enzymes	9
4.2-7 Changes of the mean-group number of cells and mean-group	
activity of SOD and CAT initiated by combined (γ +laser)	^
and (laser+y) irradiation	J
4.2-8 The effect of gamma and combined irradiation	_
on the individual number of cells	2
4.2-9 The effect of the laser energy density on the post-radiation	_
number of cells and SOD activity)
4.3 Possible mechanisms of the radioprotective action of low-intensity	7
optical radiation (laser/LED)	
4.4 References	1
Conclusions 21	5

LIST OF FIGURES

Fig. 1-1 Absorption spectra: whole blood (A1) and erythrocytes (A2); whole blood with a higher content of oxyhemoglobin (B2) and with a lower one (B1); plasma (C)
Fig. 1-2 Variation of the shape of the absorption bands at 541 and 577 nm
with increase in the content of oxyhemoglobin (F(HbO ₂))
Fig. 1-3 Infrared absorption spectra of whole blood (A) and plasma
(B)
Fig. 1-4 Deconvolution spectra of the infrared absorption bands of
erythrocytes: (A) in range of Amide A; (B) in the ranges of Amide I
and Amide II ($\Delta v_{1/2}=100 \text{ cm}^{-1}$ (A), 30 cm ⁻¹ (B) and $k_{enh.}=2.5$)
Fig. 1-5 Infrared absorption spectra of films: (1) prepared from an aqueous
solution of ceftriaxone (concentration c=10 mg/mL, dark line) and (2)
from whole blood (film thickness $d = 5 \mu m$, light line)
Fig. 1-6 Ratio of optical densities D_{bl}/D_{ceftr} (the blood bands at v_{max} = 1654
(1), 1540 (2), 1402 cm ⁻¹ (3) and ceftriaxone band at v_{max} = 1764 cm ⁻¹)
vs. ceftriaxone concentration ($r = 0.98$, $p<0.05$ (1); $r = 0.99$, $p<0.05$ (2,
3))
Fig. 1-7 (A) Plasma fluorescence spectra (λ_{ex} = 280 nm): a healthy person
(1) and a CVD patient (2); excitation spectrum (3); (B) deconvolution
spectra of the plasma fluorescence of a healthy person (1), the CVD
patient (2) ($\Delta v_{1/2} = 40 \text{ nm}$ and $k_{enh} = 2.5$) [36]
Fig. 1-8 UVBI effect on the plasma absorption spectra of 2 patients: solid
line – before UVBI; dotted line – after 5 UVBI procedures
Fig. 1-9 Electronic absorption spectra of the blood samples: (1) before
UVBI and (2) after 5 UVBI procedures with $E = 0.06 \text{ J/cm}^2$; (A) the
Soret bands normalized by intensity at the band maximum; (B) doublet
of bands at 541 and 577 nm
Fig. 1-10 (A) Electronic absorption spectra of erythrocytes: before UVBI
(1) and after 5 UVBI procedures (2); (B) the Soret bands normalized
by intensity at the band maximum; (C) doublet of bands at 541 and 577
nm (the notation as before) 22
Fig. 1-11 The Soret bands in the blood absorption spectra of two CVD
patients before UVBI (A1; B1), during the UVBI procedure (A2) and
after the end of the UVBI course (B2). The corresponding

deconvolution spectra before UVBI (C1; D1), during the UVBI procedure (C2), after the end of the UVBI course (D2)
Fig. 1-12 (A) Electronic absorption spectra of blood: before irradiation (1)
and one hour after the end of 8 OVBI procedures (2); (B) the Soret
bands normalized by intensity at the band maximum; (C) doublet of
the bands at 541 and 577 nm ($\lambda_{ir.}$ = 780 nm, the laser power 10 mW, the
magnetic field strength 35 mT, the irradiation time $t = 15$ min).
Designations (1) and (2) are the same as in A
Fig. 1-13 Infrared absorption spectra of blood before IVBI (solid line) and
after the end of the course (dotted line). (λ_{ir} = 670 nm, the exposure
time $t = 20 \text{ min}, 2 \text{ mW}$ at the output of the light guide)
Fig. 1-14 Deconvolution spectra of infrared absorption bands of whole
blood: (A) Amide I and Amide II ranges; (B) Amide III range (before
IVBI (1) and after 8 IVBI procedures (2) ($\Delta v_{1/2}$ = 30 cm ⁻¹ (A), $\Delta v_{1/2}$ =
35 cm^{-1} (B) and $k_{\text{enh.}} = 2.5$))
Fig. 1-15 Deconvolution spectra of the infrared absorption band of the NH
stretching vibrations (whole blood samples): (1) before OVBI; (2) after
OVBI ($\lambda_{ir} = 670 \text{ nm}$)
Fig. 1-16 Infrared spectra of plasma samples prepared from blood taken
before irradiation (solid line) and after the end of the IVBI course
(dotted line) ($\lambda_{ir.}$ = 632.8 nm)
Fig. 1-17 Deconvolution spectra of infrared absorption bands of
erythrocytes for 2 patients in the Amide I and II ranges: (A) increasing
and (B) decreasing the content of oxyhemoglobin: before OVBI (1)
and after OVBI (2) (λ _{ir.} =670 nm)
Fig. 1-18 Deconvolution spectra of the infrared bands of blood in the
Amide III range (normalized by the intensity at the maximum of the
band at v_{max} =1232.5 cm ⁻¹). (A) Increasing and (B) decreasing the
content of oxyhemoglobin: before OVBI (1) and after OVBI (2) at $\lambda_{ir.}$ =
670 nm
Fig. 1-19 Infrared absorption spectra of blood (A) and deconvolution
spectra (B) in the range of the band of stretching vibrations of
phosphate groups: before IVBI (1), after 8 IVBI procedures (2) ($\Delta v_{1/2}$ =
30 cm ⁻¹ , k _{enh.} = 2.5)
Fig. 1-20 Infrared absorption spectra of erythrocytes: before (solid line)
and after 5 UVBI procedures with $E = 0.07 \text{ J/cm}^2$ (dotted line) 35
Fig. 1-21 Deconvolution spectra of the infrared absorption bands of
erythrocytes: (A) the Amid A range (normalized by intensity at the
maximum of the band at 3300 cm ⁻¹); (B) the Amide I and Amide II
ranges (normalized by intensity at the maximum of the band at 1655
cm ⁻¹): solid curve – before UVBI, dashed curve – after 5 UVBI

procedures with E = 0.07 J/cm ² ($\Delta v_{1/2} = 100 \text{ cm}^{-1}$ (A) and $\Delta v_{1/2} = 30$
cm^{-1} (B), $k_{enh}=2.5$)
Fig. 1-22 Deconvolution spectra of the infrared absorption bands of
plasma: (A) the Amide A range ($\Delta v_{1/2}$ = 100 cm ⁻¹ , k_{enh} = 2.5); (B)
Amide I and Amide II ranges ($\Delta v_{1/2}$ = 30 cm ⁻¹ , $k_{enh.}$ = 2.5), solid line –
before UVBI, dotted line - after 5 UVBI procedures with E= 0.07
J/cm ²
Fig. 1-23 Infrared deconvolution spectra of the blood absorption bands in
the Amide III range (normalized by the intensity at the maximum of
the band at 1306 cm ⁻¹): before (1) and after UVBI (2), ($\Delta v_{1/2} = 30$
cm ⁻¹ , k _{enh.} = 2.5); (A) the decrease and (B) the increase of the
oxyhemoglobin content
Fig. 1-24 (A-C) Plasma fluorescence spectra of 3 CVD patients: before (1)
and after the UVBI course (2); (A'-C') the corresponding
deconvolution spectra ($\Delta \lambda_{1/2} = 40$ nm, $k_{enh.} = 2.5$) ($\lambda_{ex} = 280$ nm) 39
Fig. 1-25 Plasma fluorescence spectra of 2 patients: (A1) before IVBI
$(\lambda_{.ir.}$ = 670 nm, 3mW, the exposure time t=15 min); (B1) before OVBI
($\lambda_{ex.}$ = 670nm, 10mW, the exposure time t=15 min), (A2) after 5 IVBI
procedures and (B2) after 5 OVBI procedures; (A', B') -
corresponding deconvolution spectra ($\Delta \lambda_{1/2}$ = 35 nm, $k_{enh.}$ = 2.5) 40
Fig. 1-26 Maxima position (λ_{max}) of the plasma fluorescence spectra vs.
pH values: ● – before UVBI, O – after UVBI (r= 0.7, p<0.05)
Fig. 2-1 Changes in the gas partial pressures in venous blood of one of the
patients during the IVBI course ($\lambda = 670$ nm): (1) p_VO_2 before the
procedure (\square) and during the procedure (\blacksquare); (2) p_VCO_2 before the
procedure (O) and during the procedure (O)
Fig. 2-2 Changes in the oxygen partial pressure p _V O ₂ in venous blood of
one of the patients: (A) during and after UVBI and (B) during and after
OVBI procedure ($\lambda_{ir.} = 670 \text{ nm}$)
Fig. 2-3 Photoinduced changes in the oxygen partial pressure ($\Delta p_V O_2$) and
the carbon dioxide partial pressure ($\Delta p_V CO_2$) vs. $\Delta S_V O_2$ under UVBI (r
= 0.81, p<0.001 (A), and $r = -0.8$, p<0.004 (B)). Each point reflects the
changes of the gas pressures in blood of one patient
Fig 2-4 Photoinduced changes in the oxygen partial pressure $(\Delta p_{\rm V} O_2)$ and
the carbon dioxide partial pressure ($\Delta p_V CO_2$) vs. $\Delta S_V O_2$ under OVBI (r
= 0.81, p<0.001 (A), and $r = -0.70$, p<0.004 (B)). Each point reflects
the changes of the gas partial pressures in blood of one patient 53
Fig. 2-5 (A) The oxygen partial pressure p _V O ₂ in venous blood during the
UVBI course. (B) The changes in the growth of p_VO_2 (Δp_VO_2) during
the IVBI course. For both UVBI and IVBI, the results belong to one
patient

Fig. 2-6 Changes of the carbon dioxide partial pressure (Δp _V CO ₂) initiated
by UVBI vs. the initial values of p_VCO_2 in venous blood (r = -0.61,
p<0.01). Each point reflects the changes in blood of one patient 55
Fig. 2-7 Normalizing UVBI effect on the mean-group partial pressures
p _V CO ₂ in patient's blood of the two subgroups with different initial
values of p _V CO ₂ and, as a consequence, with a different direction of
p _V CO ₂ changes in these groups after 5 UVBI procedures
Fig. 2-8 (A) Photoinduced changes in the bicarbonate ion concentration
(ΔC_{HCO3}) vs. ΔS_VO_2 . (B) Changes in pH value (ΔpH) vs. ΔS_VO_2 in
venous blood under UVBI. (A) $r = -0.64$, p<0.002; (B) $r = 0.52$,
p<0.02). Each point in Figs 2-8 reflects the changes in blood of one
patient
Fig. 2-9 Photoinduced changes in the bicarbonate ion concentrations
(ΔC _{HCO3} ⁻) vs. initial values (C _{HCO3} ⁻) in venous blood: (A) under UVBI
(r = -0.7, p < 0.0012) and (B) under OVBI $(r = -0.66, p < 0.0015)$. Each
point in Figs 2-9 reflects the changes of the bicarbonate ion (HCO ₃ ⁻)
concentration in blood of one patient
Fig. 2-10 Normalizing effect of 5 UVBI procedures on the mean-group
concentrations of bicarbonate ion in blood of the patients of two
subgroups with different initial concentrations
Fig. 2-11 Changes of the degree of hemoglobin oxygen saturation in
venous blood ($\Delta S_V O_2$) after the end of PT course vs. initial $S_V O_2$
values: (A) after UVBI and (B) after OVBI. Each point corresponds to
the results obtained for one patient
Fig. 2-12 Changes in the degree of hemoglobin oxygen saturation $S_{V}O_{2}$ in
venous blood of one of the patients during the OVBI course: (0)
-before the procedure, (\square) -during the procedure. The numbers
indicate the procedure number
Fig. 2-13 The degree of hemoglobin oxygen saturation (S _V O ₂) in venous
blood vs. the procedure number. The S _V O ₂ values are measured after
the end of procedures: (1) IVBI and (2) UVBI. Each point reflects the
value of S _V O ₂ in venous blood of one patient
Fig. 2-14 AVDO ₂ changes vs. ΔS_VO_2 after the OVBI course (A) and after
the UVBI course (B). Each point in the Figs corresponds to the data for
one patient. (A) $r = -0.97 \text{ p} < 0.001$; (B) $r = -0.95 \text{ p} < 0.001 \dots 66$
Fig. 2-15 Oxyhemoglobin dissociation curves (ODC) before UVBI (1, 2)
and after UVBI (3, 4) for the CVD patients with different initial blood
oxygenation. Patient 1 (ODC-1, 3): before UVBI $-p_VO_2 = 19.8$ mm
Hg, $S_VO_2 = 32.3\%$; after UVBI $-p_VO_2 = 24.1$ mm Hg, $S_VO_2 = 44.2\%$;
patient 2 (ODC-2, 4): before UVBI $-p_VO_2 = 26.5$ mm Hg, $S_VO_2 =$
39.1%; after UVBI $-p_VO_2 = 20.2 \text{ mm Hg}$, $S_VO_2 = 30.9\%$

Fig. 2-16 The p50 changes (Δ p50) vs. the initial p50 value in venous blood
of individual patients: (A) after UVBI and (B) after OVBI courses. (A)
$r = -0.68 \text{ p} < 0.001$; (B) $r = -0.72 \text{ p} < 0.001 \dots 68$
Fig. 2-17 Photoinduced p50 changes (Δ p50) vs. Δ S _V O ₂ after the UVBI
course (A) ($r = 0.64$, p<0.006) and (B) after the OVBI course ($r = 0.55$,
p<0.013). Each point corresponds to the result for one patient 69
Fig. 2-18 The RBC volume changes during IVBI procedures. The values
denoted by (★) are obtained during the procedure
Fig. 2-19 Electrophoretic mobility of erythrocytes in the CVD patients 77
Fig. 2-20 Changes during the UVBI course: Hct in blood samples of
patients with different initial Hct values (1, 2); K ⁺ ion concentration
(3), and Na ⁺⁻ ion concentration (4) in blood samples of patient number
1
Fig. 2-21 Changes during the UVBI course: Hb concentration (C _{Hb}) in
blood samples of patients with high initial C _{Hb} (1) and with normal
$C_{Hb}(2)$; the oxygen partial pressure $p_VO_2(3)$ for patient 1
Fig. 2-22 Changes during the IVBI course: (1) Hb concentration, (2)
oxyhemoglobin content, and (3) K ion concentration in blood of one of
the patients
Fig. 2-23 Changes in hemoglobin concentration (ΔC_{Hb}) vs. the initial C_{Hb}
value after the UVBI (\bullet) and after OVBI (\square) courses, $r = -0.7$,
p<0.001). Each point corresponds to the data for one patient
Fig. 2-24 Changes of hemoglobin oxygen saturation in venous blood
(S _V O ₂) during PT courses. (1) The UVBI course: before the procedure
(•), (0) during procedure in blood samples taken from UV cuvette; (2)
before the UVBI procedure (A), in blood samples taken during the
procedure (\triangle); (3) the OVBI course: before procedure (\blacksquare), during the
procedure (□)
Fig. 2-25 (1, 2) The changes of oxygen utilization rate (Rutil.) in two
patients during the UVBI course; (3) oxygen partial pressure in venous
blood (p _V O ₂) in patient 1 and (4) in patient 2
Fig. 2-26 Relative mean-group blood viscosity η_{bl} , η_w before and after the
UVBI course
Fig. 2-27 Relative mean-group plasma viscosity $\eta_{pl}/\eta_{w.}$ before and after
the UVBI course86
Fig. 2-28 Changes, initiated by UVBI in the blood viscosity ($\Delta\eta_{bl.}$) (r=
-0.63 at p<0.04) and the plasma viscosity ($\Delta \eta_{pl}$) (r = -0.75 at p<0.01)
vs. their initial values in individual patients
Fig. 2-29 Changes in aPTT (Δ_{aPTT}), TC (Δ_{TC}) and PT ratio ($\Delta_{PT ratio}$)
initiated by UVBI vs. their initial values ($r = -0.64$, p<0.002 for aPTT;
r = -0.8, p<0.001 for TCT; $r = -0.57$, p<0.02 for PT ratio)
/ 1 · · · · / · · · · / · · · · · · · ·

Fig. 2-30 Regulatory effect of 5 UVBI procedures on the mean aPTT
values for two subgroups of patients having different initial values 89
Fig. 2-31 Changes in aPTT (Δ_{aPTT}) initiated by OVBI vs. aPTT values in
individual patients ($r = -0.64$, p<0.002)90
Fig. 2-32 Changes in TC (Δ_{TC}) initiated by OVBI vs. TC initial values in
individual patients ($r = -0.8$, p<0.001)
Fig. 2-33 OVBI initiated changes in PT ratio (Δ _{PT ratio}) vs. PT ratio initial
values in individual patients ($r = -0.78$, p<0.002)90
Fig. 2-34 OVBI initiated changes in INR (Δ_{INR}) vs. INR initial values in
individual patients ($r = -0.72$, p<0.001)
Fig. 2-35 Changes of TCT and aPTT under UVBI (●) and OVBI (□): (A)
Δ_{TCT} vs. $\Delta S_V O_2$; (B) aPTT (Δ_{aPTT}) vs. $\Delta S_V O_2$
Fig. 2-36 Oscillations during the IVBI course ($\lambda_{ir.}$ = 670 nm): (1) oxygen
partial pressure p_VO_2 (before the procedure (\square) , during the procedure
(■)), (2) lactate concentration C _{lac.} (before IVBI (O), during the IVBI
procedure (●))
Fig. 2-37 Changes initiated by UVBI in the glucose and lactate
concentrations: (A) ΔC_{gl} vs. C_{gl} ; (C) ΔC_{lac} vs. C_{lac} ; (B, D) ΔC_{gl} and
ΔC_{lac} vs. the S_VO_2 photoinduced changes (ΔS_VO_2) $(r$ = -0.75 for
p<0.0001 (A) and r= -0.62 for p<0.001 (C))
Fig. 2-38 (A) Changes initiated by OVBI (●) and UVBI (□) in the glucose
$(\Delta C_{gl.})$ and lactate (ΔC_{lac}) concentrations: (A) ΔC_{gl} vs. C_{gl} ; ΔC_{lac} vs.
$C_{lac};$ (B) $\Delta C_{gl.}$ and $\Delta C_{lac.}$ vs. ΔS_VO_2- the photoinduced changes in S_VO_2
(r= -0.58 for p<0.01 (ΔC_{gl}) and r= -0.41 for p<0.06 ($\Delta C_{lac.}$)). Each
point corresponds to the data for one patient
Fig. 2-39 Concentration changes initiated by UVBI in cholesterol fraction
in the different patients: (A) ΔC_C vs. C_C (r= -0.69, p<0.01), (B)
ΔC_{LDL-C} vs. C_{LDL-C} (r = -0.70, p<0.01), (C) ΔC_{HDL-C} vs. C_{HDL-C} (r =
0.82, p<0.01), (D) ΔC_{TG} vs. C_{TG} (r = -0.58, p<0.01). Each point in Fig.
2-39 corresponds to the data for one patient
Fig. 2-40 Changes initiated by UVBI in the concentrations of total
cholesterol (C _C) and low density lipoprotein cholesterol (C _{LDL-C}) in
patients with the different degree of hemoglobin oxygen saturation:
(A) ΔC_{TC} vs. $\Delta S_V O_2$; (B) ΔC_{LDL-C} vs. $\Delta S_V O_2$. Each point in Fig. 2-40
corresponds to the data for one patient
Fig. 2-41 UVBI-initiated change in infrared plasma absorption bands in
the range of symmetric CH-stretching vibrations for 2 patients: (A) the
cholesterol concentration in the first patient was C _C =4.8 mmol/L
before UVBI (curve 1) and 4.1 mmol/L after UVBI (curve 2); (B) in

the second patient – C_C = 7.9 mmol/L before UVBI (curve 1) and 5.9
mmol/L after UVBI (curve 2)101
Fig. 2-42 Deconvolution spectra of infrared plasma absorption bands in
the ranges of Amide I and Amide II: the patient with C_C =7.9 mmol/L
before UVBI (solid line) and 5.9 mmol/L after UVBI (dotted line) 101
Fig. 2-43 Normalizing effect of 5 UVBI procedures on cholesterol
concentration in two patient subgroups which differ the initial
cholesterol concentrations and, as a consequence, the direction of
changes in cholesterol concentration (C _C) after the UVBI course (■-
concentration decrease, — concentration increase)
Fig. 2-44 Changes of relative erythrocyte concentrations (ΔC _{RBC} /C _{RBC})
under UVBI in patients with different cholesterol concentrations:
$\Delta C_{RBC}/C_{RBC}$ vs. $\Delta C_{C}/C_{C}$
Fig. 2-45 Effect of the initial C _C and C _{C-LDL} concentrations in blood of the
individual patients on the change in their concentrations initiated by
OVBI (r= -0.70 , p<0.01 for ΔC_C vs. C_C and r= -0.58 , p<0.01 for
ΔC _{LDL-C} vs. C _{LDL-C})
Fig. 2-46 The OVBI effect on mean-group lipid concentrations in a group of
CVD patients (n=25). Before the course: (1) C, (3) LDL-C, (5) HDL-C,
(7) TG and after the course end – (2), (4), (6), (8) respectively
Fig. 2-47 Changes in LDL-C concentration (ΔC_{LDL-C}) in individual
patients vs. $\Delta S_V O_2$ -photoinduced changes in the degree of hemoglobin
oxygen saturation in venous blood (ΔS _V O ₂)109
Fig. 3-1 Absorption spectra: (1) blood (hematocrit (Ht)-44%), (2) water
[24]. Phosphate buffer: pH = 7.4; 0.3 mol/L
Fig. 3-2 Spectral dependences of the scattering coefficient μ_s and the
anisotropy factor g for blood [26]. Phosphate buffer: pH = 7.4; 0.3
mol/L
Fig. 3-3 Spectral dependences of the skin coefficients: absorption – μ_a and
scattering – μ_s , measured in vivo [33]
Fig. 3-4 (A) The blood transmission vs. z – the light penetration depth for
different wavelengths: (1) λ =524 nm; (2) λ =630 nm; (3) λ =905 nm); skin
tissue transmission: (4) λ =524 nm; (5) λ =630 nm; (6) λ =890 nm. (B) The
transmission spectrum of skin tissues at depths of 1 and 2 mm
Fig. 3-5 A volume distribution of the intensity of the optical radiation
sources in a vein of cylindrical form in the plane of section, which is
normal to the axis of the cylinder
Fig. 3-6(A)The depth of light penetration into blood and (B) into the skin
tissue vs. wavelength of optical radiation: (1) the reduced scattering

coefficient; (2) the spectrum of the effective penetration depth; (3) the
spectrum of full penetration depth
Fig. 3-7 The $\mu_a(\lambda)/\mu_{ef}(\lambda)$ ratio as a function of wavelength. The effective
spectral absorption coefficients of blood ($C_{ef}(\lambda, z)$) are shown by
vertical bars
Fig. 3-8 The dependence of the OVBI induced changes on the initial
degree of the hemoglobin oxygen saturation S _V O ₂ (curve 2 of each
panel): (A) oxyHb content ($r = -0.89$, p<0.001); (B) glucose
concentration (r= -0.54 , p<0.001); (C) the concentration of cholesterol
(r = -0.67, p < 0.001). Curves 1 of each panel show the dependence of
the blood absorption coefficient μ_a on S_VO_2
Fig. 3-9 Regulatory effects of 7 OVBI procedures: (A) the degree of
hemoglobin oxygen saturation S _V O ₂ ; (B) the cholesterol concentration
C _C : (●) for the patients of the subgroup with values higher than median
value; (I) for the patients of the subgroup with values lower than
median value
Fig. 3-10 (A) The temperature of the tissue heating (T) under OVBI as a
function of the irradiation time t: (1) blood in the blood vessel (λ_{ir} =
780 nm, $I_0 = 0.02 \text{ W/cm}^2$; (2) on the front wall of the vessel; (3) on the
skin surface; (B) heating temperature T vs. the tissue penetration depth
z: (1) $\lambda_{ir} = 470$ nm, $I_0 = 0.035$ W/cm ² ; (2) $\lambda_{ir} = 670$ nm, $I_0 = 0.25$
W/cm^2 ; (3) $\lambda_{ir} = 780 \text{ nm}$, $I_0 = 0.01 \text{ W/cm}^2$
Fig. 3-11 A volume temperature distribution under OVBI ($\lambda_{ir.}$ = 780 nm,
0.004 W/cm^2 , $t = 10 \text{ min}$) in a vein d= 3 mm. The arrow indicates the
direction of the radiation propagation
Fig. 3-12 The temperature of tissue heating under OVBI ($\lambda_{ir.}$ = 670 nm, I_0 =
0.25 W/cm ²) vs. the irradiation time t: (1) on the skin surface; (2) on
the front wall of the vessel; (3) blood in the blood vessel 144
Fig. 3-13 The temperature of tissue heating under OVBI ($\lambda_{ir.}$ = 470 nm, I_0 =
0.035 W/cm ²) vs. irradiation time t: (1) on the skin surface; (2) on the
front wall of the vessel; (3) blood in the blood vessel
Fig. 3-14 (A) Deconvolution spectra of the Soret bands ($\Delta \lambda_{1/2}$ = 50 nm and
k_{enh} = 2.5) of blood samples: (1) intact rats; (2) samples taken on the
second (3) and on the 5th day after OVBI. (B) Absorption spectra of
blood samples in the range of 600–1000 nm: (1) intact rats, samples
taken on the 2nd (2) and (3) on the 5th day after OVBI
Fig. 3-15 Temperature of tissue heating (T) under OVBI (λ_{ir} =2000 nm) vs.
irradiation time t: (A) on the skin surface (dotted line) and in the blood
vessel (solid lines); (B) temperature variation over depth of the tissue
z. Radiation power densities: 1 W/cm ² (1), 2 W/cm ² (2), and 6.4
W/cm^2 (3)

Fig. 3-16 Distribution of the MB concentration over the depth of the
mucosal tissue for tissue staining time $t_{\text{stain}}=1$ (1), 2.5 (2), 5 (3) and 10
(4) minutes. (5) Relative sensibilizer amount S _{rel} diffusing into a layer
d=500 μm of mucosal tissue vs. staining time
Fig. 3-17 Distribution of laser energy, absorbed per unit volume of
scattering tissue in the direction of propagation of the laser radiation
$(Q,W/cm^3)$, $\lambda_{ir}=670$ nm, $I=0.09$ W/cm ² . (A) (1) without the dye and
(2) for thickness of the stained layer d= 50 μ m, C(t) _{rel} = 0.32, t _{stain} = 10
min, μ_{PS} = 4.5 cm ⁻¹ . (B) (1) without the dye; (2) for thickness of the
stained layer d = 500 μ m, C(t) _{rel} = 0.43, t _{stain} = 2.5 min, μ _{PS} = 25.9 cm ⁻¹ ;
(3) $C(t)_{rel} = 0.56$, $t_{stain} = 5$ min, $\mu_{PS} = 34.0$ cm ⁻¹ ; (4) $C(t)_{rel} = 0.715$, $t_{stain} = 0.715$
10 min, μ_{PS} = 43.0 cm ⁻¹
Fig. 3-18 Distribution of laser energy absorbed per unit volume of
scattering tissue in the direction of the radiation propagation
$(Q,W/cm^3)$, $\lambda_{ir.}$ = 632.8 nm, I= 0.0105 W/cm ² . (A) (1) without the dye;
(2) for thickness of the stained layer d= $500\mu m$, $C(t)_{rel} = 0.715$, $t_{stain} =$
10 min, μ_{PS} = 2.5 cm ⁻¹ ; (B) (3) C(t) _{rel} = 0.29, t _{stain} = 1.0 min, μ_{PS} = 4.64
cm ⁻¹ ; (4) C(t) _{rel} = 0.43, t_{stain} = 2.5 min, μ_{PS} = 6.91 cm ⁻¹ ; (5) C(t) _{rel} =
0.56 , $t_{\text{stain}} = 5.0 \text{ min}$, $\mu_{\text{PS}} = 8.96 \text{ cm}^{-1}$
Fig. 3-19 Temperature distribution over the cross section of the mucosal
tissue layers: (A) (1) when irradiated with laser radiation at λ_{ir} = 670
nm, intensity I= 0.09 W/cm ² without the dye and (2) for thickness of
the stained layer d= 500 μ m, C(t) _{rel} = 0.43, t _{stain} = 2.5 min, μ _{PS} = 25.9
cm ⁻¹ (2); $C(t)_{rel} = 0.56$, $t_{stain} = 5$ min, $\mu_{PS} = 34.0$ cm ⁻¹ (3); $C(t)_{rel} = 0.715$,
t_{stain} = 10 min, μ_{PS} = 43.0 cm ⁻¹ (4); (B) (1) when irradiated with laser
radiation at $\lambda_{ir} = 632.8$ nm, intensity I= 0.0105 W/cm ² without the dye
and (2) for thickness of the stained layer d= 500 μ m, C(t) _{rel} = 0.29,
$t_{\text{stain}} = 1.0 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 0.43, \ t_{\text{stain}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 2.5 \text{ min}, \ \mu_{PS} = 4.64 \text{ cm}^{-1}; \ (3) \ C(t)_{\text{rel}} = 2.5 \text{ min}, \ \mu_{PS} = $
6.91 cm^{-1} ; (4) C(t) _{rel} = 0.56, t _{stain} = 5.0 min, μ_{PS} = 8.96 cm ⁻¹
Fig. 3-20 Infrared absorption spectra of blood samples: before the
magnetic therapy (1) and after one procedure (2)
Fig. 3-21 Infrared absorption spectra of blood samples before (1) and after
the magnetic therapy (2): for Amide I band (A), Amide III band (B).
Deconvolution spectra of the corresponding bands before (3) and after
the magnetic therapy (4) ($\Delta v_{1/2} = 30 \text{ cm}^{-1}$, $k_{\text{enh}} = 2.5$)
Fig. 3-22 Absorption spectra of blood samples in the range of 1000–1150
cm ⁻¹ : before (1) and after the magnetic therapy (2). Deconvolution
spectrum of the blood sample after the magnetic therapy (3) ($\Delta v_{1/2}$ = 30
cm^{-1} , $k_{enh} = 2.5$)
Fig. 3-23 The ROS formation in the processes of single-electron reduction
and excitation of oxygen

Fig. 3-24 Changes of the oxygen partial pressure $(\Delta p_V O_2)$ and
arteriovenous oxygen difference (Δ_{AVD}) vs. photoinduced changes
S_VO_2 (ΔS_VO_2), obtained after the end of UVBI (\bullet) and OVBI courses
(\square) ($\lambda_{ir.}$ = 670 nm). For $\Delta p_V O_2$ vs. $\Delta S_V O_2$: $r=0.67$, $p<0.001$ and for
Δ_{AVD} vs. $\Delta S_V O_2$; $r = -0.56$, $p < 0.001$
Fig. 3-25 Changes in the cholesterol concentration (ΔC_{C}) and APTT
values (Δ_{APTT}) after the end of UVBI (\bullet) and OVBI courses (\Box) $(\lambda_{ir.} =$
670 nm) as functions of their initial values. For ΔC_C vs. C_C , $r = -0.67$,
p<0.001; for Δ_{APTT} vs. APTT – r = -0,56, p<0.001
Fig. 4-1 The transmission spectra of the rat tissues: layers with thicknesses
of 1 mm (1), 1.5 mm (2), and 2 mm (3)
Fig. 4-2 The absorption spectra of the venous blood samples in rats (№ 3-
5) in the range of 300-700 nm: (1) before OVBI and (2) after OVBI
$(E=7.5 \text{ J/cm}^2)$
Fig. 4-3 The absorption spectra of venous blood samples in rats (№ 8–10)
in the range of 650–1100 nm: (1) before irradiation; (2) after (laser+ γ)
irradiation (E= 7.5 J/cm ²); (3) after γ -irradiation (3Gy)
Fig. 4-4 Infrared absorption spectra of the venous blood samples of rats:
$№ 8.2$ and $№ 10.2$ after (laser+ γ) irradiation, № 8.3 and № 10.3 after γ -
irradiation
Fig. 4-5 Changes in the mean-group number of WBC and LYM under
OVBI at E = 7.5 J/cm ² and single γ -irradiation (3 Gy) in series of
experiments (A) and (B), differing the mean-group initial cell number.
WBC: (A) (1) control group, (2) single γ-irradiation; (3) 3 OVBI
procedures followed by γ -irradiation; (B) (1) control group, (2) 3
OVBI procedures; (3) single γ -irradiation; (4) 3 OVBI procedures
followed by γ -irradiation. LYM%: (A) (1) control, (2) γ -irradiation; (3)
3 OVBI procedures followed by γ -irradiation; (4) γ -irradiation
followed by 3 OVBI procedures; (B) (1) control, (2) γ irradiation; (3) 3
OVBI procedures followed by γ -irradiation, (4) 3 OVBI procedures.
(C) The relative changes in the mean-group number of C_{WBC} and C_{LYM}
caused by γ -radiation (3 Gy) vs. initial concentrations (C _{WBC} (C) and
$C_{LYM}(C)$
Fig. 4-6 Changes in the mean-group RBC and Hb concentrations in series
I with the most powerful changes. (A) RBC: (1) control group, (2) 3
OVBI procedures followed by γ -irradiation, (3) γ -irradiation (3 Gy),
(4) γ -irradiation followed by 1 OVBI procedure (E = 6.25 J/cm ²). Hb:
(1) control group, (2) 3 preliminary OVBI procedures (E = 7.5 J/cm^2),
(3) γ -irradiation (3 Gy), (4) 3 OVBI procedures (E = 7.5 J/cm ²)
followed by γ-irradiation

Fig. 4-7 Changes in PLT mean-group concentration at the laser energy density $E = 5 \text{ J/cm}^2$ and γ -radiation (3 Gy) in series of experiments (A) and (B), differing the mean-group initial values. (A): (1) control group; (2) γ-irradiation; (3) 4 OVBI procedures followed by γ-irradiation; (4) 4 OVBI procedures; (B): (1) control group; (2) γ- irradiation; (3) 4 OVBI procedures followed by γ -irradiation; (4) single γ -irradiation followed by 4 OVBI procedures; (5) 4 OVBI procedures. (C) The relative changes in the mean-group number of CPLT caused by yradiation (3 Gy) as a function of the C_{PLT} initial concentration in Fig. 4-8 Changes in the mean-group activity of SOD in blood samples of rats in two series of experiments (A) and (B), differing in the meangroup initial SOD activity. (A): (1) control group, (2) γ-irradiation (3Gy); (3) 3 OVBI procedures (E = 7.5 J/cm^2) followed by γ irradiation; B: (1) control group, (2) γ-irradiation; (3) 4 OVBI procedures (E = 5 J/cm²) followed by γ -irradiation; (4) preliminary γ irradiation followed by 4 OVBI procedures (E = 5 J/cm²). C: The relative decrease in activity of SOD caused by γ -radiation as a function Fig. 4-9 The relative number of cells: $(C_{LIM} (laser + \gamma)/C_{LYM} (\gamma) (\bullet); C_{WBC}$ (laser $+\gamma$)/C_{WBC} (γ) (\square); LYM % (laser $+\gamma$)/LYM %(γ) (\blacktriangle)) vs. the relative values of the SOD activity obtained at the same conditions of Fig. 4-10 The changes in the individual number of leukocytes (ΔC_{WBC}) and lymphocytes (ΔC_{LYM}) as function of the initial values C_{WBC} (C) or C_{LYM} (C) in individual rats. (A): (1) γ -irradiation (3 Gy) (\bullet); γ irradiation and 3 followed OVBI procedures of at $E = 7.5 \text{ J/cm}^2$ (O); 4 OVBI procedures at E = 5 J/cm² followed by γ -irradiation (\blacktriangle) (r = -0.5, p<0.001); (2) 4 OVBI procedures at E = 5 J/cm² (r= -0.85, p<0.004) (\bigstar); (B): γ -irradiation (3 Gy) (\square); γ -irradiation followed by 3 OVBI procedures at E = 18.75 J/cm² (\triangle); γ - irradiation followed by 3 Fig. 4-11 Individual changes in ΔC_{Hb} and ΔC_{RBC} vs. initial values C_{Hb} and C_{RBC} in different rats. (A): γ-irradiation (3 Gy) (•); 3 preliminary OVBI procedures (E = 7.5 J/cm²) followed by γ -irradiation (\blacksquare); 3 OVBI procedures (E = 7.5 J/cm²) (O); (r = -0.68, p <0.001). (B): yirradiation (\square); 3 OVBI procedures (E = 7.5 J/cm²) followed by γ -Fig. 4-12 (A) Individual changes in C_{PLT} : ΔC_{PLT} vs. C_{PLT} (C) $(E_{OVBI} = 5$ J/cm²) (1) γ-irradiation (**1**); γ-irradiation followed by 3 OVBI procedures at $C_{PLT}(C) = 944 \cdot 10^9 / L$; (r=-0.93, p<0.001) (\bullet); (2) y-

irradiation (\square), 4 OVBI procedures followed by γ -irradiation (\circ); γ -
irradiation followed by 4 OVBI procedures (\triangle) at $C_{PLT}(C)$ =
$708 \cdot 10^9$ /L; (r = -0.86, p<0.002). (B) Individual changes in SOD
activity: 3 OVBI procedure (E = 7.5 J/cm ²) followed by γ -irradiation (
•); γ -irradiation followed by 3 OVBI procedures (E= 7.5 J/cm ²) (\triangle); 4
OVBI procedures (E = 5 J/cm ²) followed by γ -irradiation (\square) (r =
-0.85, p<0.0001)
Fig. 4-13 The relative number of cells and SOD activity as a function of
the total laser energy density E used at combined irradiation: several
preliminary OVBI procedures followed by γ -irradiation (\bullet);
preliminary γ -irradiation and then several OVBI procedures (\square) 206

LIST OF TABLES

Table 1-1 UVBI effect: on the maximum position of the plasma fluorescence spectra (λ ^{fl} _{max}) and their component (λ ^{fl} _{max} (comp); on the acid-based parameters (pH, p _v CO ₂ , HCO ⁻ ₃), measured on an ABL-800 instrument. The values of pH, pCO ₂ and HCO ⁻ are presented taking into account the coefficients of a variation for ABL-800 instrument
Table 2-1 Mean-group blood oxygenation values and the acid-based status
of blood in CVD patients, the oxygen capacity of venous blood ct_VO_2
and arterial blood ct _A O ₂ , the arteriovenous difference in the oxygen
content AVDO2 before the PT course and one day after the course,
p50–the partial pressure of O_2 at which $S_VO_2=50$ %
Table 3-1 Characteristics of the human tissues (λ _{ir} .=780 nm)
Table 3-2 Characteristics of the human tissues (λ_{ir} .= 670 nm)
Table 3-3 Characteristics of the human tissues (λ_{ir} = 2000 nm)
Table 3-4 Optical and thermal characteristics of mucosal tissue
Table 4-I Relative changes (γ/control) in the mean-group RBC and Hb
concentrations, Hct values in various series of experiments 195
Table 4-2 Radioprotective effect of laser radiation on the main parameters
of peripheral blood and the activity of antioxidant enzymes under γ -
irradiation of the whole body of rats (3 Gy) and different variants of
OVBI

ACKNOWLEDGEMENTS

We are very grateful to our colleague-scientists, whose help made a contribution to the writing of this monograph:

Professor Ulastchik V.S.-Academician of the National Academy of Sciences of Belarus, Professor, D. Sc. (Med), a scientific editor of the first edition of this monograph in 2014; many thanks for valuable advices and comments:

Professors, D. Sc. (Med.): Nechipurenko N.I., Kirkovski V.V., Marochkov A.V. – Competent consultants on the use of UV and intravenous blood irradiation in medical practice;

Professors, D. Sc. (Phys. & Math.): Asimov M.M, Zeltov G.I., Bushuk B.A., Dzagarov B.M., many thanks for fruitful discussion of photophysical processes initiated by low-intensity optical radiation in human tissues;

Sambor E.G., Kuchinskii A. V., Maslova T.O., Galay O.A., Karoza A.V., Batay L.E.—Colleagues from the B.I. Stepanov Institute of the Physics of the National Academy of Sciences of Belarus, conducted experiments and Astaf'eva L.G. for invaluable assistance in modeling the optical radiation propagation in human tissues; special thanks to Kalosha I.I. for both many-year fruitful cooperation and invaluable help in discussing and preparing the monograph;

Laskina O.V. – Assistant of the 3rd Department of Internal Medicine at the Belarussian State Medical University; whose enthusiasm and dedicated work arrived at the unique results on the individual sensitivity of patients to low-intensity optical radiation effects;

Nasek V.M., Zilberrman R.G. – Colleagues from the Institute of Bioorganic Chemistry of the National Academy of Sciences of Belarus and Koshlan I,V. – Colleague from the Joint Institute for Nuclear Research (Dubna, Russia) for assistance in conducting experiments on the combined effects of gamma and laser irradiations on rats.

We are very much obliged for your help.

INTRODUCTION

The optical radiation effect on blood in blood vessels and tissues, used for therapeutic purposes, is one of the methods of a phototherapy (PT) that is the fastest-growing field of modern medicine. Laser medicine originated in the second half of the twentieth century; it should be viewed as a PT subset that is one of the oldest therapeutic methods. PT was initially utilized in medical practice as solar therapy, then light therapy, and then as ultraviolet blood irradiation that has been in use in medicine for 70 years. Currently, PT performed as intravenous and overvein blood irradiation with low-intensity optical radiation is considered as a method of biostimulation suitable for treating a wide range of diseases. Evidence has emerged that the combined effects of phototherapy and drug treatment improve treatment outcomes.

So far, the expansion of the scope of applications of intravenous and overvein blood irradiation has been empirical. The widespread use of these methods in medical practice was limited by a lack of generally accepted ideas about the primary molecular mechanism of the optical radiation action on blood irradiated in vivo. The previously proposed biostimulation mechanisms, based on the analysis of various secondary reaction products, obtained by irradiating cell cultures and biotissues, remained at the level of the hypotheses unable to explain many experimental facts. In addition, it was impossible to predict the PT effects on a living organism by the processes detected in cell cultures and in isolated tissues. It appeared that a direct comparison of the effects observed in model experiments and in a living organism poses great difficulties. The same PT methods, with identical energy densities of light exposure, initiated improvements in some patients and unpredictable negative reactions in others. The positive and negative treatment results were not explained scientifically. Such topical photobiostimulation problems remained unresolved, as the development of methods for assessing their effectiveness for individual patients and the creation of methods for standardization and control.

In this regard, the study of photoreactions initiated in blood by therapeutic doses of the optical radiation at different wavelengths, the determination of laws of blood photomodification, and the identification of factors that most affect the individual patient sensitivity to the blood irradiation are among the urgent tasks of PT. Solving these problems will contribute to deepening our understanding of PT mechanisms and also further expand the applications of laser-optical technologies in medicine.

ABOUT THE STRUCTURE OF THE BOOK

The book is founded on the experimental results of the authors. The main purpose of this book is to propose and substantiate PT molecular mechanisms with the implication of photophysical processes initiated by low-intensity optical radiation. The book is organized in four chapters, a conclusion and a bibliography.

According to the basic law of photochemistry, a photochemical reaction in blood can occur only if irradiated blood is capable of absorbing incident light. In Chapter One, we studied the spectral characteristics of whole blood, red cells and plasma before and after UV extracorporeal, intravenous and overvein blood irradiation by low-intensity laser-optical radiation at different wavelengths. The spectral manifestations of photochemical processes initiated in blood under PT were explored and explaind in order to examine the irradiation effects at the molecular level. Fluorescent spectroscopy was adopted to investigate possible structural changes in plasma macromolecules after the blood irradiation *in vivo*.

In Chapter Two, the research objects were blood gas composition, a degree of hemoglobin oxygen saturation in venous and arterial blood, acid-based balance indices, erythrocyte and hemoglobin concentrations, as well as contents of some metabolic products under PT. A study of changes initiated by PT in the partial pressure of blood gases and in the degree of hemoglobin oxygen saturation is a further step in identifying the photoacceptor molecule. Based on these data, the blood irradiation effect on the oxygen status of venous and arterial blood, the oxygen supply to the organs and tissues, and the oxygen utilization was studied. The blood oxygenation dynamics during the PT courses was defined as well. The changes in hematocrit, hemoglobin concentration, and erythrocyte count at low-intensity optical radiation doses were monitored during the PT course. Special attention was given to the susceptibility of individual patients to blood irradiation, and methods for its control were suggested.

A significant part of this chapter was devoted to metabolic processes under PT. Using glucose and lactate (playing an important role in metabolic processes), as an example we studied the PT effect on their concentrations. We showed that therapeutic doses of the optical radiation absorbed by blood cause short- and long-term changes in lactate and glucose concentrations in blood of patients. The dependences of

concentration changes initiated by PT on both the initial concentrations and the photoinduced variations in the blood oxygenation were analyzed.

Using the optical radiation of various wavelengths, we investigated the PT effects on blood coagulation parameters in different patients. The concentration changes in total cholesterol, triglycerides, low-density lipoprotein cholesterol, and high-density lipoprotein cholesterol were estimated. A relationship between the photoinduced changes in blood oxygenation characteristics and the concentrations of metabolic products was analyzed. Possible molecular mechanisms were considered for the effects observed.

Chapter Three was devoted to analyzing the molecular mechanism of the PT action. First, a brief review based on the analysis of literature data was presented. It covered the proposed mechanisms of therapeutic action of laser-optical radiation. It should be noted that before our studies, the PT effect on a living organism had not been sufficiently analyzed at the molecular level.

The hypothesis that local heating of blood or tissues in the absorption zone is a starting mechanism for biostimulation has been actively discussed to date. We therefore performed a computer simulation of thermal fields and estimated heating temperatures at the tissue surface, in tissues, and in venous blood at low- and high-intensity laser irradiation. Computer modeling of low-intensity optical radiation effects in mucosal tissues was used to explore the mechanism of the optical radiation action on tissues stained by photosensitizers.

The choice of a wavelength for PT is another medical problem, for the solution of which the optical properties of blood and tissues are extremely important. The optical properties of blood and human tissues were briefly summarized. Based on these data, spectra of the radiation penetration depth into blood and skin tissues were calculated in the wavelength range of 405–950 nm. The wavelengths optimal for PT were then determined. Molecular oxygen is effectively formed during intravenous and overvein blood irradiation with the optical radiation at these wavelengths.

Based on the results obtained, we proposed a molecular mechanism for the therapeutic action capable of explaining a number of effects that had not been interpreted unambiguously for many years. The main photoactivation stages of the organism were suggested. The possible role of reactive oxygen species in the therapeutic action of low-intensity optical radiation was considered. The applicability of the proposed PT mechanism was discussed to explain the therapeutic effects observed in treating a wide range of diseases.

In Chapter Four, which deals with rat experiments, the radioprotective possibilities of low-intensity optical radiation were used to study the actual problem of counteracting the destructive action of ionizing radiation. The available literature data have no convincing confirmation of the radioprotective capability of low-intensity optical radiation. The effect of *in vivo* blood irradiation with the laser radiation on the hematological parameters and the antioxidant protection enzymes for gamma-exposed rats was examined. The quantitative changes caused by low-intensity laser radiation and gamma radiation in blood cells were analyzed. Differences in the individual radiosensitivity of rats were investigated. The molecular mechanisms of the radioprotective effect of low-intensity optical radiation, as well as the factors affecting the individual sensitivity of rats were discussed.

In conclusion, the results obtained by the authors of the book were summarized. The most important results were emphasized. In our opinion, they will contribute to a further successful use of PT in medical practice. Some important issues, which remained unresolved despite a rapid growth of a number of PT studies, were considered. Further research areas are outlined.

The book summarizes the interdisciplinary research results, and will be of interest to researchers in photophysics, biophysics, physiotherapy, as well as to physicians using phototherapy in complex treatment and rehabilitation of patients. The book is also intended as a teaching tool either for graduate students or students interested in physical medicine problems.

ABBREVIATIONS

ACS – Acute coronary sindrome AOS – Antioxidant system

aPPT – Activated partially prothromboplastin time

AVDO₂ – Arteriovenous oxygen difference

Ca²⁺ – Calcium ions CO₂ – Carbon dioxide

ConO₂ – Consumption of oxygen CVD – Cardiovascular disease

CAT – Catalase

C_{util} – Coefficient of oxygen utilization ct_aO₂ – Oxygen capacity in arterial blood ct_VO₂ – Oxygen capacity in venous blood

DRA – Deoxyribonucleic acid 2,3-DFG – 2,3-Diphosphoglycerate DO₂ – Delivery of oxygen

EFME – Electrophoretic mobility of erythrocyte

EMT – Extracorporeal magnetotherapy F_V(HbO₂) – Content of oxyhemoglobin fraction

GRA – Granulocyte Gy – Gray

Hct - Hematocrit

Hb - Hemoglobin

H⁺ - Hydrogen ions

H₂O₂ - Hydrogen peroxide

HbO₂ - Oxyhemoglobin

HCO₃ - Hydrocarbonate ion

IR - Ionizing radiation

INR – International normalized ratio
 IVBI – Intravenous blood irradiation
 k_{enh}. – Resolution enhancement factor

KRS – Thallium bromide iodide mixed crystal

LED – Light emitting diode

LILR – Low intensity laser radiation LIOR – Low intensity optical radiation xxviii Abbreviations

LPO – Lipid peroxidation LYM – Lymphocyte

LTM - Lymphocyte

MCV – Mean erythrocyte volume

MetHb – Methemoglobin MF – Magnetic field MT – Magnetic therapy NO – Nitrogen oxide

ODC – oxyhemoglobin dissociation curve OTFB – Oxygen transport function of blood

OVBI – Overvein blood irradiation

O₂ Oxygen

¹O₂ – Singlet oxygen

O₂⁻ - Superoxide anion radical PMM - Primary molecular mechanism

PPA – Primary photoacceptor

pH - Blood acidity

P₅₀ – Oxygen partial pressure at which hemoglobin is 50%

saturated

Δp_VO₂ – Oxygen partial pressure in venous blood

pvCO₂ – Carbon dioxide partial pressure in venous blood

r – Pearson linear correlation coefficient

RBC – Erythrocyte RNA – Ribonucleic acid

ROS – Reactive oxygen species
SOD – Superoxide dismutase
TCT – Trombin cloting time

UV – Ultraviolet

UVBI – Ultraviolet blood irradiation

S_VO₂ – Hemoglobin oxygen saturation in venous blood

V_{er} – Volume of red blood cell

WBC - Leukocyte

 $\begin{array}{cccc} \gamma & & - & Gamma\ radiation \\ \eta_{bl} & & - & Blood\ viscosity \\ \eta_{pl} & & - & Plasma\ viscosity \\ \lambda & & - & Wavelength \end{array}$

 $\Delta v_{1/2}$ — Half width of absorption band

CHAPTER ONE

THE EFFECT OF LOW-INTENSITY OPTICAL RADIATION ON THE SPECTRAL-LUMINESCENT CHARACTERISTICS OF BLOOD IRRADIATED IN VIVO

1.1 Electronic absorption spectra of whole blood, erythrocytes and plasma

Absorption spectroscopy is widely used to study the physicochemical properties of biological objects [1–3]. The position of absorption bands, and their half-width and intensity provide information on a qualitative and quantitative ratio of the components of biological systems and on their structural organization in the ground electronic state.

In the wavelength range of 210–250 nm, a number of blood components are absorbed: residues of aliphatic amino acids of proteins, free aliphatic amino acids of blood plasma, lipids of a membrane and plasma, polysaccharides, and some other non-protein organic components of blood plasma. In the range of 250–300 nm, the absorption occurs due to residues of aromatic amino acids: tryptophan, tyrosine, and phenylalanine. Of these, tryptophan has the highest absorption coefficient at a wavelength of 254 nm which is used in UV devices for the blood irradiation. Free amino acids and radicals of blood plasma that play an important role in initiating photooxidation reactions of unsaturated fatty acids are also absorbed within this range. It should be noted that UV radiation at 254 nm used for therapeutic purposes is photochemically active and can cause a breakdown of nucleic acids, proteins and lipids.

The absorption spectra of whole blood, erythrocytes and plasma are shown in Fig. 1-1. The shortwave absorption band, with a maximum wavelength of λ_{max} = 280 nm, is present in the spectra of all proteins containing aromatic amino acid residues.

The absorption bands of deoxyhemoglobin at $\lambda_{max} = 430$ and 555 nm, as well as the oxyhemoglobin bands at $\lambda_{max} = 344$, 415, 541 and 577 nm,

which appear in the blood and erythrocyte spectra, are caused by the absorption of iron porphyrin being part of hemoglobin [4]. Hemoglobin is a protein globule-tetramer. It is formed by four polypeptide chains, each of which is an individual molecule containing a protoporphyrin chelate complex with iron (heme), capable of attaching ligands (O₂, CO, NO, etc.). In the UV range, the absorption bands at λ_{max} =274 nm for deoxyhemoglobin and λ_{max} =276 nm for oxyhemoglobin, corresponding to their shortest wave transition, are masked by the absorption of proteins.

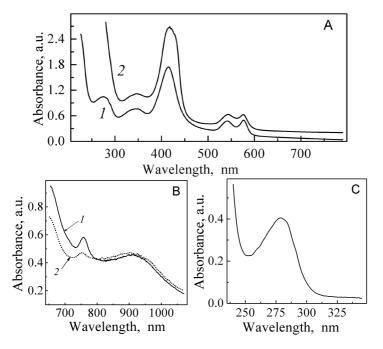


Fig. 1-1 Absorption spectra: whole blood (A1) and erythrocytes (A2), whole blood with a higher concentration of oxyhemoglobin (B2) and with a lower one (B1); plasma (C)

In the absorption spectrum of all hemoglobins, the most intense among iron-porphyrin bands is the Soret band ($k=140 \text{ mM}^{-1} \cdot \text{cm}^{-1}$), the maximum of which can be between $410 < \lambda_{\text{max}} < 430 \text{ nm}$. According to the data [5], in the range of the Soret band, the electronic absorption spectra of porphyrins have two resolved high-intensity bands which belong to transitions, polarized mutually perpendicularly. High-frequency asymmetry of the band contour is due to the vibrational structure. The assignment of the